



## PAPER

## The effect of short and continuous absorbent patch application on local skin temperature underneath

RECEIVED  
7 January 2021REVISED  
19 March 2021ACCEPTED FOR PUBLICATION  
30 March 2021PUBLISHED  
12 May 2021Lisa Klous , Mireille Folkerts , Hein Daanen  and Nicola Gerrett 

Department of Human Movement Sciences, Faculty of Behavioural and Movement Sciences, Vrije Universiteit Amsterdam, Amsterdam Movement Sciences, Amsterdam, The Netherlands

E-mail: [h.a.m.daanen@vu.nl](mailto:h.a.m.daanen@vu.nl)**Keywords:** absorbent patch; sweat rate; skin temperature; local**Abstract**

**Objective.** By attaching absorbent patches to the skin to collect sweat, an increase in local skin temperature ( $T_{sk}$ ) underneath the patches seems unavoidable. Yet this effect has not been quantified. The present study investigates the effect of absorbent patch application on local  $T_{sk}$  underneath.

**Approach.** Ten healthy participants cycled for 60 min at an exercise intensity relative to their body surface area ( $40 \text{ W}\cdot\text{m}^{-2}$ ) in three environmental conditions (temperate:  $25^\circ\text{C}$  45% RH, hot-humid:  $33^\circ\text{C}$  65% RH and hot-dry:  $40^\circ\text{C}$  30% RH). The effect of short sweat sampling (i.e. from min 25–30 to min 55–60) on  $T_{sk}$  was examined on the right scapula.  $T_{sk}$  of the left scapula served as control. The effect of continuous sweat sampling (i.e. four consecutive 15 min periods) on  $T_{sk}$  was examined on the right upper arm.  $T_{sk}$  of the left upper arm served as control. **Main results.** Neither short nor continuous application of absorbent sweat patches affected  $T_{sk}$  underneath the patches in the hot-humid and hot-dry condition ( $P > 0.05$ ). In the temperate condition, continuous application led to a significant increase in  $T_{sk}$  underneath the patches during the first and second minute. This increase remained throughout the experiment ( $1.8 \pm 0.6^\circ\text{C}$ ;  $P < 0.001$ ). Short application of sweat patches did not affect the local  $T_{sk}$  underneath ( $P > 0.05$ ) in the temperate condition. **Significance.** To avoid a significant increase in local  $T_{sk}$  underneath sweat patches, continuous application should be prevented in, especially, a temperate condition. Timely removal of sweat patches should be taken into account during longer periods of collecting sweat in field or laboratories settings.

**1. Introduction**

Measuring local sweat rate (LSR) has become popular in research laboratories to understand sweat gland function, in sports to quantify electrolyte losses, and in industry for functional clothing design and designing thermal and sweating manikins. A convenient method to determine LSR and sweat content is the absorbent patch technique (Smith and Havenith 2011, 2012, Baker *et al* 2016, 2018, Klous *et al* 2020a, 2020b).

For the determination of LSR, Smith and Havenith proposed short sampling durations ( $<5$  min) to prevent hidromeiosis (i.e. reduction of sweating by blockage of the sweat glands due to humid conditions) (Brown and Sargent 1965, Candas *et al* 1984) underneath the patches. Secondly, sweat sampling was recommended once a steady sweat rate has been achieved. In their studies, therefore, usually sweat was sampled for the final 5 min of a 30 min exercise period, from now on referred to as ‘short sweat sampling’ (Smith and Havenith 2011, 2012). However, to assess sweat composition a certain sweat volume is required to be able to perform the chemical analysis. Since commonly used analyzers for sweat have a relatively large dead volume, large volumes of sweat are required to be able to perform the intended chemical analysis (for example 0.3 ml). Based on experience from our lab, a sampling period of  $\sim 15$  min is required to collect enough sweat for chemical analysis of more than two sweat components. These 15 min sampling periods are typically seen consecutively (Klous *et al* 2020a, 2020b), which is from now on referred to as ‘continuous sweat sampling’.

Covering the skin with absorbent material and impermeable material on top to prevent sweat loss creates a microclimate which could elevate local skin temperature ( $T_{sk}$ ) (Brebner and Kerslake 1964, Psikuta *et al* 2014, MacRae *et al* 2018). Utilizing both sweat sampling methods, an increase in local  $T_{sk}$  underneath patches seems unavoidable, which may affect LSR. Previous research reported an obvious effect of the change in  $T_{sk}$  on sweating (Kondo *et al* 1997, Wurster and McCook 1969, Libert *et al* 1979). The onset threshold for sweating was also reached earlier when having a higher  $T_{sk}$ , whereas the threshold occurred later with a lower  $T_{sk}$  (Nadel *et al* 1971a, 1971b, Cotter and Taylor 2005, Shibasaki *et al* 2006). Furthermore, eccrine sweat glands have a so-called thermal sensitivity: local heating increases and local cooling decreases LSR for a given core temperature (Nadel *et al* 1971a, 1971b, Cotter and Taylor 2005, Shibasaki *et al* 2006). This may be mediated by an increased neurotransmitter release for a given neural stimulus in case of local heating and desensitization in case of cooling. However, scientific evidence to back up this theory is lacking. Recently, Gerrett *et al* (2019) showed that the reabsorption rates of eccrine sweat glands were higher, once  $T_{sk}$  increased considerably, which may in turn affect sweat content. Yet the effect of applying absorbent patches on  $T_{sk}$  has to be quantified.

The present study investigates the effect of short and continuous application of absorbent sweat patches on local  $T_{sk}$  underneath the patches. This was done in three types of environmental conditions that are representative for sweat-related research: temperate (25 °C, 45% RH), hot-humid (33 °C, 65% RH), and hot-dry (40 °C, 30% RH). It was hypothesized that both short and continuous application of absorbent sweat patches affect  $T_{sk}$ , but that the increase in  $T_{sk}$  underneath the patches would be statistically non-significant in any of the three environmental conditions.

## 2. Material and methods

### 2.1. Ethical approval

Procedures were approved by the Ethics Committee of the Faculty of Behavioural and Movement Sciences of the Vrije Universiteit Amsterdam (VCWE-2020-142; amendment VCWE-2020-171). The study was conducted in accordance with the guidelines of the revised *Declaration of Helsinki* (2013). Written informed consent was obtained from all participants before participation in the study.

### 2.2. Participants

Ten healthy individuals (4 males, 6 females; age:  $29 \pm 4$  years; height:  $175.6 \pm 8.2$  cm; weight:  $71.3 \pm 12.5$  kg) participated in this study. Participants were instructed to refrain from alcohol 24 h before the experiment, to limit caffeine consumption and to consume 500 ml of water 0–2 h before starting the experiment. No further restrictions were placed on participants diets. All participants were non-smokers, did not take any prescription medication, had no history of heat-related illnesses, cardiovascular complications and did not have any known issues with thermoregulation. Menstrual cycle phase was not controlled for in the female participants.

### 2.3. Design

Participants visited the laboratory three times to cycle (Lode Excalibur, Groningen, The Netherlands) for 60 min at an exercise intensity relative to their body surface area, corresponding to  $40 \text{ W}\cdot\text{m}^{-2}$  (Cramer and Jay 2014), in a climate chamber (b-Cat, Tiel, The Netherlands) set to either 25 °C 45% RH, 33 °C 65% RH or 40 °C 30% RH. Airflow in the climate chamber was standardized to  $0.2 \text{ ms}^{-1}$  for all environmental conditions. The order of tests was balanced. To account for changes due to circadian rhythms, each visit was scheduled at the same time of day ( $\pm 3$  h). The effect of short sweat sampling (i.e. from min 25–30 to min 55–60; figure 1) on  $T_{sk}$  was examined on the right scapula.  $T_{sk}$  of the left scapula served as control. Likewise, the effect of continuous sweat sampling (i.e. four consecutive 15 min periods; figure 1) on  $T_{sk}$  was examined on the right upper arm.  $T_{sk}$  of the left upper arm served as control.

### 2.4. Measurements

To confirm hydration, urine specific gravity (USG) was measured with a handheld refractometer (PAL-S, Atago, Bellevue, USA;  $\text{USG} \leq 1.025$ ) (Kenefick and Chevront 2012). Despite the hydration guidelines, one participant had an  $\text{USG} > 1.025$  in the temperate condition and drunk  $5 \text{ ml}\cdot\text{kg}^{-1}$  of water before resuming the experiment. Temperature sensors (Thermochron DS1922L, Maxim Integrated, USA) were attached to the left and right scapula (zone 13 as reported by Gerrett *et al* 2014) and upper arm (zone 19 as reported by Gerrett *et al* 2014). The  $T_{sk}$  sensor on the right arm was continuously covered (i.e. four consecutive sampling periods of 15 min) by absorbent material (Cutisoft, BSN Medical, Almere, The Netherlands, size:  $25 \text{ cm}^2$  ( $5 \times 5$  cm), absorbent capacity:  $\sim 2$ g). The sweat collection period of 15 min was chosen because that time period is the minimum to allow for collection of enough sweat for chemical analysis and is short enough to prevent saturation of the patch. The absorbent material was covered by an impermeable layer (Parafilm-M, Bemis, Saint Louis,



**Table 1.** Mean (SD) control skin temperatures of short and continuous sweat sampling in three environmental conditions ( $n = 10$ ).

Environmental condition	Control $T_{sk}$ -short sampling ( $^{\circ}\text{C}$ )	Control $T_{sk}$ -continuous sampling ( $^{\circ}\text{C}$ )
Temperate (25 $^{\circ}\text{C}$ 45% RH)	32.7 (1.0)	33.1 (1.0)
Hot-humid (33 $^{\circ}\text{C}$ 65% RH)	36.2 (1.0) <sup>a</sup>	36.5 (1.0) <sup>a</sup>
Hot-dry (40 $^{\circ}\text{C}$ 30% RH)	36.7 (1.0) <sup>a,b</sup>	36.8 (1.1) <sup>a,b</sup>

Note.  $T_{sk}$ : skin temperature; RH: relative humidity.

<sup>a</sup>  $P < 0.05$  from temperate.

<sup>b</sup>  $P < 0.05$  from hot-humid.

USA, size: 30.25 cm<sup>2</sup> (5.5 × 5.5 cm)) and was attached to the skin by a porous adhesive (Fixomull stretch, BSN Medical, Almere, The Netherlands, size: 100 cm<sup>2</sup> (10 × 10 cm)). The  $T_{sk}$  sensor on the right scapula was covered by the same sweat patch from min 25–30 to min 55–60 on the arm (i.e. short sweat sampling) as proposed by Smith and Havenith (2011, 2012). The corresponding left skin sites, arm and scapula, served as control.  $T_{sk}$  sensors on the control sites were only covered by a 25 cm<sup>2</sup> (5 × 5 cm) piece of the porous adhesive Fixomull that is commonly used in practice to attach sensors to the skin.

## 2.5. Statistics

$T_{sk}$  data were recorded and presented every minute. Statistical analysis were performed using IBM SPSS Statistics 26.0. Effects were considered significant if  $P < 0.05$ . Data are presented as means and standard deviations (SD). The Shapiro-Wilk test was used to check if the data was normally distributed. To assess the effect of environmental condition on control  $T_{sk}$ , a one-way ANOVA was conducted employing the independent variable environmental condition (temperate, hot-humid and hot-dry). This was done separately for the short and continuous sweat sampling method. Secondly, the effect of sweat sampling over time on  $T_{sk}$  was analyzed using a two-way ANOVA (sampling × time) for both sampling methods (short and continuous sampling) and three environmental conditions (temperate, hot-humid and hot-dry) separately. If there were significant effects, specific post-hoc analyses were conducted with the alpha adjusted accordingly using Bonferroni corrections.

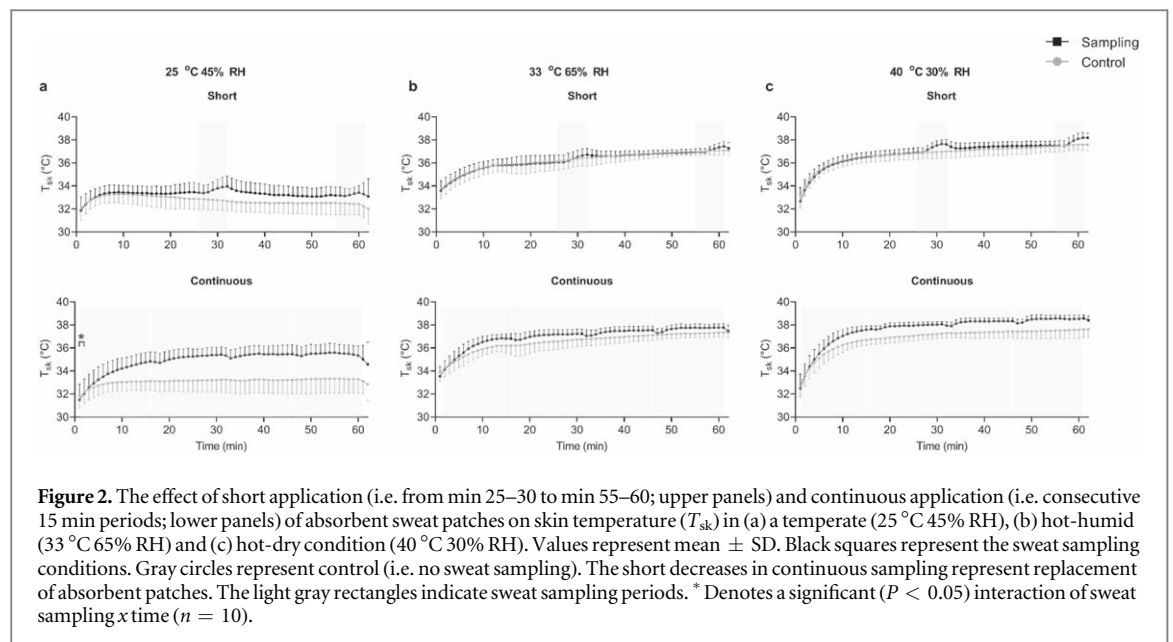
## 3. Results

### 3.1. The effect of environmental condition on control $T_{sk}$

There was a significant main effect of environmental condition on control  $T_{sk}$  during both short and continuous sampling methods ( $P < 0.001$ ; table 1). Post-hoc analysis revealed that for both sampling methods, control  $T_{sk}$  was significantly higher in the hot-dry and hot-humid compared to the temperate condition ( $P < 0.001$ ; table 1). For both sampling methods, control  $T_{sk}$  was also significantly higher in the hot-dry than the hot-humid condition ( $P < 0.001$ ; table 1).

### 3.2. The effect of short and continuous sweat sampling on $T_{sk}$

The sampling × time interaction was non-significant in the hot-humid and hot-dry condition for both sampling methods ( $P \geq 0.177$ ; figure 2). In the temperate condition, there was a significant sampling × time interaction due to continuous sampling ( $P < 0.001$ ), but not due to short sampling ( $P = 0.845$ ). Post-hoc analysis revealed that the sampling × time interaction was significant from min 1–2 ( $P \leq 0.047$ ; figure 2). This indicates that due to continuous sampling the  $T_{sk}$  pattern increased more from min 1–2 compared to control. After that, sampling and control  $T_{sk}$  patterns over time were similar ( $P \geq 0.072$ ), indicating that the initially elevated  $T_{sk}$  remained throughout the protocol.



## 4. Discussion

Sweat absorbent patches are a convenient sweat sampling technique for both LSR and sweat content but their impact on local microclimate  $T_{sk}$  in different conditions is not well known. This research is important because large differences in local  $T_{sk}$  can modulate the sweat response. The present study showed that neither short (i.e. from min 25–30 to min 55–60) nor continuous application (i.e. four consecutive 15 min periods) of absorbent sweat patches significantly affected  $T_{sk}$  underneath the patches in two environmental conditions (hot-humid: 33 °C 65% RH and hot-dry: 40 °C 30% RH). In a temperate condition (25 °C 45% RH), short application of sweat patches had no effect either, but continuous application led to an immediate significant increase in  $T_{sk}$  ( $1.8 \pm 0.6$  °C) underneath the patches. This elevated  $T_{sk}$  remained throughout the experiment.

### 4.1. Effect of absorbent sweat patch application

In the present study, there was no effect of short and continuous application of absorbent sweat patches on  $T_{sk}$  in the hot-humid condition. During heat stress, the most prominent autonomic (i.e. involuntary) thermoregulatory responses are vasodilation and evaporation of sweat. Vasodilation increases  $T_{sk}$  by internal conductive heat exchange between tissue and blood, which in turn increases convective and radiant heat loss from the skin to the environment. Convective heat loss through vasodilation is a function of the air velocity over the body surface area. In addition, convective and radiant heat loss are a function of the temperature gradient between the air and skin. If the environmental temperature is high, the temperature gradient between air and skin is small, which decreases the effectiveness of vasodilation. Therefore, in hot environments the evaporation of sweat is the most prominent autonomic thermoregulatory response (Taylor 2014, Periard *et al* 2015, Cramer and Jay 2016). The driving force for evaporation of sweat is the difference in vapor pressure between the skin and air (Parsons 2014, Cramer and Jay 2016). In a hot-humid environment, evaporation of sweat is difficult due to the high water vapor content of the air (Parsons 2014, Cramer and Jay 2016). Covering the skin with absorbent material with an impermeable layer on top also inhibits the ability to evaporate sweat. Since this heat loss mechanism is already suppressed due to the environment, the added effect of the application of sweat patches may be negligible, explaining our finding.

In the hot-dry condition, there was again a statistically non-significant  $T_{sk}$  elevation over time by applying sweat patches. In dry heat, the vapor pressure difference between skin and air is relatively large, allowing for evaporation of sweat. Due to the high environmental temperature (40 °C) compared to  $T_{sk}$  ( $\sim 36.7$  °C), there was most likely no heat loss but rather heat gain through convection and radiation (assuming radiant temperature was identical to air temperature) (Parsons 2014, Cramer and Jay 2016). Suppressing the evaporation of sweat, on which the body heavily relies in the hot-dry condition, could explain the slightly larger elevations in  $T_{sk}$  compared to the hot-humid condition. However, this effect did not reach statistical significance either. Because there was dry heat gain already, adding the absorbent patch probably did not contribute to any further significant increase in  $T_{sk}$ .

In contrast, there was an effect of the continuous application of absorbent sweat patches on  $T_{sk}$  in the temperate condition. In this condition, the temperature gradient between air and skin was relatively large,

allowing for convective and radiant heat loss (Parsons 2014, Cramer and Jay 2016). In addition, the vapor pressure gradient between air and skin was also relatively large, allowing for evaporation of sweat. In terms of heat loss from the human body, this condition was considered most beneficial. Thus, in the most beneficial condition, the  $T_{sk}$  effect of prolonged absorbent patch application on  $T_{sk}$  is considerable. As mentioned in the introduction, this significantly higher  $T_{sk}$  could affect the processes of sweating (Nadel *et al* 1971a, 1971b, Cotter and Taylor 2005, Shibasaki *et al* 2006, Gerrett *et al* 2019) and should therefore be avoided.

#### 4.2. Minimal relevant difference

During single absorbent patch applications, elevations in  $T_{sk}$  ranged from  $0.05\text{ }^{\circ}\text{C} \pm 0.1\text{ }^{\circ}\text{C}$  to  $0.8\text{ }^{\circ}\text{C} \pm 0.1\text{ }^{\circ}\text{C}$  using the short method (i.e. sampling for 5 min) and from  $0.4\text{ }^{\circ}\text{C} \pm 0.3\text{ }^{\circ}\text{C}$  to  $2.2\text{ }^{\circ}\text{C} \pm 0.1\text{ }^{\circ}\text{C}$  using the continuous method (i.e. sampling for 15 min). The smallest elevations were observed in the hot-humid condition and the largest elevations in the temperate condition. In relation to this, Nadel *et al* (1971a), (1971b) showed that when maintaining a constant esophageal temperature, a  $1\text{ }^{\circ}\text{C}$  higher whole-body  $T_{sk}$  (from  $34.0\text{ }^{\circ}\text{C}$  to  $35.0\text{ }^{\circ}\text{C}$  and from  $35.0\text{ }^{\circ}\text{C}$  to  $36.0\text{ }^{\circ}\text{C}$ ) corresponded to a  $\sim 0.18\text{ mg}\cdot\text{cm}^{-2}\cdot\text{min}^{-1}$  higher LSR at the thigh. This was reported in the presence of a whole-body  $T_{sk}$  elevation, whilst the local  $T_{sk}$  underneath their ventilated capsule remained  $34.8\text{ }^{\circ}\text{C}$  throughout. In the present study, the  $T_{sk}$  manipulation was only local. To our knowledge, the effect of a local  $T_{sk}$  elevation on LSR is not quantified. Yet previous research suggests that a higher local  $T_{sk}$  affects neurotransmitter release at the neuroglandular junction (Ogawa 1970, Ogawa and Asayama 1986, DiPasquale *et al* 2003), causing a higher LSR. The largest mean elevation in  $T_{sk}$  ( $2.2\text{ }^{\circ}\text{C} \pm 0.1\text{ }^{\circ}\text{C}$ ; figure 1) was observed using the continuous method (fourth patch) in the temperate condition. Previous research by Gerrett *et al* (2019) reported higher reabsorption rates in the eccrine sweat gland if  $T_{sk}$  increased by more than  $6\text{ }^{\circ}\text{C}$ . The discrepancy between these values is large and therefore the eccrine sweat glands reabsorption rates are not expected to be affected by the continuous absorbent patch application. Since the sweat glands reabsorption rate determines the final sweat sodium and chloride concentrations, sweat composition is not expected to be affected by short nor continuous absorbent sweat patch application. This has important applications for field work or laboratory studies investigating sweat content.

#### 4.3. Practical implication

The proposed method by Smith and Havenith to gravimetrically determine LSR using absorbent patch application for 5 min (Smith and Havenith 2011, 2012) generally does not allow for collection of enough sweat for chemical analysis. This relatively short application time is recommended to allow for a steady sweat rate and to prevent hidromeiosis underneath the patches, which could affect the sweat response (Brown and Sargent 1965, Candas *et al* 1984). For sweat content research, Baker and colleagues typically applied the absorbent patches upon moderate sweat absorption ( $\sim 0.5\text{ g}$ ) from different body regions. Application times ranged from  $\sim 15\text{ min}$  at the forehead to  $\sim 48\text{ min}$  at the calf, and they removed the patches before saturation (Baker *et al* 2009, 2014, 2016, 2019). During sweat testing in the field, longer sampling periods up to 60 min were reported due to practical reasons (Alvear-Ordenes *et al* 2005, Stofan *et al* 2005, Henkin *et al* 2010). In laboratory settings, absorbent patches were left on the skin for  $\sim 15\text{--}30\text{ min}$  (Schwartz and Thaysen 1956, Lobeck and Huebner 1962, Verde *et al* 1982, Goulet *et al* 2017, Baker *et al* 2018, Klous *et al* 2020a, 2020b). However, Morris *et al* (2013) explicitly recommend to limit patch application time to 5 min to mitigate the effect of a microenvironment on LSR. Based on experience from our lab, slightly longer consecutive sampling periods of  $\sim 15\text{ min}$  are required to collect enough sweat from the upper arm and back for duplicate chemical analysis of more than two components (Klous *et al* 2020a, 2020b). The scapula and upper arm were chosen respectively because of the relatively high LSR (Smith and Havenith 2011, 2012) and preferred location for wearables (Gao *et al* 2016). As was shown in the present study, only continuous sweat sampling (i.e. four continuous 15 min sampling periods) in the temperate condition ( $25\text{ }^{\circ}\text{C}$  45% RH) caused a significantly elevated  $T_{sk}$  underneath the patches (largest increase:  $2.2\text{ }^{\circ}\text{C} \pm 0.1\text{ }^{\circ}\text{C}$ ). To avoid any detrimental effects of absorbent sweat patch application on  $T_{sk}$ , continuous sweat sampling should not be performed in temperate conditions. To summarize, timely removal of sweat patches should be taken into account during long periods of collecting sweat in field or laboratories settings.

#### 4.4. Limitations

A limitation to the present study could be that female menstrual cycle phase was not controlled. It is known that core temperature is regulated  $\sim 0.4\text{ }^{\circ}\text{C}$  higher in the luteal compared to the follicular phase of the menstrual cycle (Kolka and Stephenson 1993). Few studies have examined the effect of menstrual cycle phase on  $T_{sk}$  and thus far results are equivocal. Part of the studies found an increased  $T_{sk}$  in the luteal phase, whilst other observed no differences relative to the follicular phase (Marsh and Jenkins 2002, Baker *et al* 2020). In the luteal phase, the onset of sweating is delayed, yet vasodilatation is increased. The net result of these processes may be a negligible

change in  $T_{sk}$  caused by menstrual cycle phase. Secondly, since short sweat sampling was investigated on the scapula and continuous sampling on the arm, there could have been an effect of skin site on our outcomes. However, since the mean differences in control  $T_{sk}$  between skin sites were very minimal (reaching from 0.1 °C to 0.4 °C; table 1), an effect of skin site is not expected. Furthermore, the effect of absorbent patch application occurred relatively fast (after 2 min of sampling) using the continuous sampling method in the temperate condition. Logically, this difference was expected to occur after at least 5 min of sampling because there were no significant differences found using the short sampling method in the same environmental condition. However, continuous sweat sampling started immediately at the onset of exercise whilst the first short sweat sampling period was 25 min into exercise (figure 1) during which we might expect to see an elevated  $T_{sk}$ . It is known that ~5–10 min are required for sweating to occur (Cotter *et al* 1997) and sweating lowers  $T_{sk}$ . Since most likely sweating (i.e. cooling the skin) did not occur during the largest part of the first continuous sweat sampling period but vasodilation did occur (i.e. heating up the skin), the fast increase in  $T_{sk}$  could be explained. However, this is true for all three conditions but only in the temperate conditions a significant  $T_{sk}$  increase was observed. Conductive and radiative heat loss through vasodilation are already limited or even heat gain occurs in the hot-humid and hot-dry condition respectively. In the temperate condition, the potential for conductive and radiative heat loss was the largest, apparently causing the largest effect of absorbents on  $T_{sk}$ . It would be interesting if future research allows for steady-state before starting a sweat sampling procedure.

## 5. Conclusions

Neither short (i.e. from min 25–30 to min 55–60) nor continuous application (i.e. four consecutive 15 min periods) of absorbent sweat patches affected skin temperature underneath the patches in a hot-humid (33 °C 65% RH) and hot-dry condition (40 °C 30% RH). In a temperate condition (25 °C 45% RH), short application of sweat patches did neither, but continuous application led to an elevated local skin temperature. To avoid a significant increase in local skin temperature underneath sweat patches, continuous application should be prevented in a temperate condition. Such findings should be taken into account when collecting sweat in field or laboratories settings.

## Acknowledgments

We would like to thank the participants who dedicated their time to this research.

## Funding

This research was supported by Eurostars under grant number 17213; and by The Dutch Research Council under grant number NWO 690462.

## Conflict of interest

The authors declare that they have no conflict of interest.

## ORCID iDs

Lisa Klous  <https://orcid.org/0000-0002-9458-4830>

Mireille Folkerts  <https://orcid.org/0000-0003-4029-6822>

Hein Daanen  <https://orcid.org/0000-0002-7459-0678>

Nicola Gerrett  <https://orcid.org/0000-0002-5210-902X>

## References

- Alvear-Ordenes I, Garcia-Lopez D, De Paz J A and Gonzalez-Gallego J 2005 Sweat lactate, ammonia, and urea in rugby players *Int. J. Sports Med.* **26** 632–7
- Baker F C, Sibozza F and Fuller A 2020 Temperature regulation in women: effects of the menstrual cycle *Temperature* **7** 226–62
- Baker L B, Barnes K A, Anderson M L, Passe D H and Stofan J R 2016 Normative data for regional sweat sodium concentration and whole-body sweating rate in athletes *J. Sports Sci.* **34** 358–68
- Baker L B, De Chavez P J D, Ungaro C T, Sopena B C, Nuccio R P, Reimel A J and Barnes K A 2019 Exercise intensity effects on total sweat electrolyte losses and regional vs. whole-body sweat [Na(+)], [Cl(-)], and [K(+)] *Eur. J. Appl. Physiol.* **119** 361–75

- Baker L B, Stofan J R, Hamilton A A and Horswill C A 2009 Comparison of regional patch collection vs. whole body washdown for measuring sweat sodium and potassium loss during exercise *J. Appl. Physiol.* **107** 887–95
- Baker L B, Ungaro C T, Barnes K A, Nuccio R P, Reimel A J and Stofan J R 2014 Validity and reliability of a field technique for sweat Na<sup>+</sup> and K<sup>+</sup> analysis during exercise in a hot-humid environment *Physiol. Rep.* **2** e12007
- Baker L B, Ungaro C T, Sopena B C, Nuccio R P, Reimel A J, Carter J M, Stofan J R and Barnes K A 2018 Body map of regional versus whole body sweating rate and sweat electrolyte concentrations in men and women during moderate exercise-heat stress *J. Appl. Physiol.* **124** 1304–18
- Brebner D F and Kerslake D M 1964 The time course of the decline in sweating produced by wetting the skin *J. Physiol.* **175** 295–302
- Brown W K and Sargent F H 1965 *Arch. Environ. Health: Int. J.* **11** 442–53
- Candas V, Brandenberger G, Lutz-Bucher B, Follenius M and Libert J P 1984 Endocrine concomitants of sweating and sweat depression *Eur. J. Appl. Physiol. Occup. Physiol.* **52** 225–9
- Cotter J D, Patterson M J and Taylor N A 1997 Sweat distribution before and after repeated heat exposure *Eur. J. Appl. Physiol. Occup. Physiol.* **76** 181–6
- Cotter J D and Taylor N A 2005 The distribution of cutaneous sudomotor and alliesthesial thermosensitivity in mildly heat-stressed humans: an open-loop approach *J. Physiol.* **565** 335–45
- Cramer M N and Jay O 2014 Selecting the correct exercise intensity for unbiased comparisons of thermoregulatory responses between groups of different mass and surface area *J. Appl. Physiol.* **116** 1123–32
- Cramer M N and Jay O 2016 Biophysical aspects of human thermoregulation during heat stress *Auton. Neurosci.* **196** 3–13
- DiPasquale D M, Buono M J and Kolkhorst F W 2003 Effect of skin temperature on the cholinergic sensitivity of the human eccrine sweat gland *Japan. J. Physiol.* **53** 427–30
- Gao W et al 2016 Fully integrated wearable sensor arrays for multiplexed *in situ* perspiration analysis *Nature* **529** 509–14
- Gerrett N, Amano T, Havenith G, Inoue Y and Kondo N 2019 The influence of local skin temperature on the sweat glands maximum ion reabsorption rate *Eur. J. Appl. Physiol.* **119** 685–95
- Gerrett N, Ouzzahra Y, Coleby S, Hobbs S, Redortier B, Voelcker T and Havenith G 2014 Thermal sensitivity to warmth during rest and exercise: a sex comparison *Eur. J. Appl. Physiol.* **114** 1451–62
- Goulet E D B, Asselin A, Gosselin J and Baker L B 2017 Measurement of sodium concentration in sweat samples: comparison of 5 analytical techniques *Appl. Physiol. Nutr. Metab.* **42** 861–8
- Henkin S D, Sehl P L and Meyer F 2010 Sweat rate and electrolyte concentration in swimmers, runners, and nonathletes *Int. J. Sports Physiol. Perform.* **5** 359–66
- Kenefick R W and Chevront S N 2012 Hydration for recreational sport and physical activity *Nutr. Rev.* **70** (Suppl. 2) S137–42
- Klous L, De Ruiter C, Alkemade P, Daanen H and Gerrett N 2020a Sweat rate and sweat composition during heat acclimation *J. Therm. Biol.* **93** 102697
- Klous L, de Ruiter C, Alkemade P, Daanen H and Gerrett N 2020b Sweat rate and sweat composition following active or passive heat re-acclimation: a pilot study *Temperature* **8** 90–104
- Kolka M A and Stephenson L A 1993 Thermoregulation in women *Exerc Sport Sci Rev.* **21** 231–62
- Kondo N, Nakadome M, Zhang K, Shiojiri T, Shibasaki M, Hirata K and Iwata A 1997 The effect of change in skin temperature due to evaporative cooling on sweating response during exercise *Int. J. Biometeorol.* **40** 99–102
- Libert J P, Candas V and Vogt J J 1979 Effect of rate of change in skin temperature on local sweating rate *J. Appl. Physiol. Respir. Environ. Exerc. Physiol.* **47** 306–11
- Lobeck C C and Huebner D 1962 Effect of age, sex, and cystic fibrosis on the sodium and potassium content of human sweat *Pediatrics* **30** 172–9
- MacRae B A, Rossi R M, Psikuta A, Spengler C M and Annaheim S 2018 Contact skin temperature measurements and associated effects of obstructing local sweat evaporation during mild exercise-induced heat stress *Physiol. Meas.* **39** 075003
- Marsh S A and Jenkins D G 2002 Physiological responses to the menstrual cycle: implications for the development of heat illness in female athletes *Sports Med.* **32** 601–14
- Morris N B, Cramer M N, Hodder S G, Havenith G and Jay O 2013 A comparison between the technical absorbent and ventilated capsule methods for measuring local sweat rate *J. Appl. Physiol.* **114** 816–23
- Nadel E R, Bullard R W and Stolwijk J A 1971a Importance of skin temperature in the regulation of sweating *J. Appl. Physiol.* **31** 80–7
- Nadel E R, Mitchell J W, Saltin B and Stolwijk J A 1971b Peripheral modifications to the central drive for sweating *J. Appl. Physiol.* **31** 828–33
- Ogawa T 1970 Local effect of skin temperature on threshold concentration of sudorific agents *J. Appl. Physiol.* **28** 18–22
- Ogawa T and Asayama M 1986 Quantitative analysis of the local effect of skin temperature on sweating *Japan. J. Physiol.* **36** 417–22
- Parsons K 2014 *Human Thermal Environments* ed K C Parsons 3rd ed (London: Taylor and Francis)
- Periard J D, Racinais S and Sawka M N 2015 Adaptations and mechanisms of human heat acclimation: applications for competitive athletes and sports *Scand J. Med. Sci. Sports* **25** (Suppl. 1) 20–38
- Psikuta A, Niedermann R and Rossi R M 2014 Effect of ambient temperature and attachment method on surface temperature measurements *Int. J. Biometeorol.* **58** 877–85
- Schwartz I L and Thaysen J H 1956 Excretion of sodium and potassium in human sweat *J. Clin. Invest.* **35** 114–20
- Shibasaki M, Wilson T E and Crandall C G 2006 Neural control and mechanisms of eccrine sweating during heat stress and exercise *J. Appl. Physiol.* **100** 1692–701
- Smith C J and Havenith G 2011 Body mapping of sweating patterns in male athletes in mild exercise-induced hyperthermia *Eur. J. Appl. Physiol.* **111** 1391–404
- Smith C J and Havenith G 2012 Body mapping of sweating patterns in athletes: a sex comparison *Med. Sci. Sports Exerc.* **44** 2350–61
- Stofan J R, Zachwieja J J, Horswill C A, Murray R, Anderson S A and Eichner E R 2005 Sweat and sodium losses in NCAA football players: a precursor to heat cramps? *Int. J. Sport Nutr. Exerc. Metab.* **15** 641–52
- Taylor N A 2014 Human heat adaptation *Compr. Physiol.* **4** 325–65
- Verde T, Shephard R J, Corey P and Moore R 1982 Sweat composition in exercise and in heat *J. Appl. Physiol. Respir. Environ. Exerc. Physiol.* **53** 1540–5
- Wurster R D and McCook R D 1969 Influence of rate of change in skin temperature on sweating *J. Appl. Physiol.* **27** 237–40